

Journal of Pharmaceutical Research International

**33(58A): 377-390, 2021; Article no.JPRI.79892 ISSN: 2456-9119** (Past name: British Journal of Pharmaceutical Research, Past ISSN: 2231-2919, NLM ID: 101631759)

### A Review of Lactoferrin Inflammatory Role in Type 2 Diabetes Mellitus with Neutrophil Dysfunction

Amani Y. Alhalwani a\*

<sup>a</sup> King Abdullah International Medical Research Center / King Saud Bin Abdulaziz University for Health Sciences, Jeddah, Saudi Arabia.

Author's contribution

The sole author designed, analysed, interpreted and prepared the manuscript.

#### Article Information

DOI: 10.9734/JPRI/2021/v33i58A34129

**Open Peer Review History:** 

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/79892

Review Article

#### Received 10 October 2021 Accepted 14 December 2021 Published 15 December 2021

#### ABSTRACT

Lactoferrin (LF) is a protein that plays important roles in many diseases including diabetes mellitus (DM). DM is one of the most challenging health concerns of the 21<sup>st</sup> century. At least 30% of the diabetic population is undiagnosed at any one time, so effective and early diagnosis is of critical concern. Several of the body's chemicals, such as enzymes, electrolytes, and proteins, have been used as biomarkers in the diagnosis of diabetic diseases. Detection of LF is considered an important sign of type 2 diabetes (T2DM), due to its activity as an anti-inflammatory agent and in the down-regulation of pro-inflammation. LF is produced by glandular epithelial cells and neutrophils, and a decrease in its concentration is linked with the dysfunction of neutrophils in many diseases. Neutrophils are the first line of defence against pathogens that invade the human body during inflammation. Therefore, the health of neutrophils can be employed as a biomarker in the diagnosis of diseases such as diabetes. A decrease in LF concentrations in T2DM could result in increased levels of inflammatory markers that are associated with the inflammation activity. Increased understanding of the link between LF concentration and development of T2DM should improve early diagnosis and treatment outcomes.

LF is identified through use of various techniques such as immunoassay, proteomics, and spectrometry. The aim of this review is to summarise each pathway and some of the most relevant LF biomarkers that may be used to monitor the development or progression of diabetes and its complications, and the link between levels of LF and neutrophil dysfunction in T2DM. Moreover, the objective of this review is to show the most common LF analysis that may be useful in the clinical diagnosis of T2DM and discuss to what extent this analysis method can be a tool for prognostic and diagnostic work.

\*Corresponding author: E-mail: amanialhalwani@gmail.com;

Keywords: Lactoferrin; Type 2 diabetes mellitus; inflammation; neutrophils; biomarkers; ELISA.

#### **1. INTRODUCTION**

Lactoferrin (LF, also known as lactotransferrin) is a protein that is produced and released by glandular epithelial cells and is detected in neutrophil secondary granules. LF is a functional glycoprotein with an estimated molecular weight of 80kDa and 690 amino acid residues [1,2]. It is found at high levels in human and bovine milk, and in smaller amounts in exocrine secretions (such as saliva, tears, sperm, vaginal fluids and gastrointestinal fluids) and cells (i.e. neutrophils, enterocytes and adipocytes) [3]. LF is a member of the glycoprotein family and has multifunctional properties. It plays an important role in the immune defence systems of the vaginal, stomach and ocular mucosa. When inflammatory stimuli are present, LF expression is enhanced in those areas, and this enhancement limits inflammatory cytokine production and the ability of lipopolysaccharide endotoxins to bind to inflammatory cells [4].

During infection, neutrophil secondary granules release increased amounts of LF at inflammatory sites to control the physiological homeostasis state [5]. LF is important in the physiological system and is used as a biomarker for many inflammatory diseases, including type 2 diabetes (T2DM). T2DM, also known as insulinindependent diabetes, is linked to obesity and insulin resistance (IR) in the peripheral tissues [6]. T2DM begins to develop several years before it is diagnosed; according to the global guideline International of the Diabetes Federation. between 30% and 90% of T2DM patients are undiagnosed at any one time [7,8]. Improved understanding of the mechanism of action of T2DM will aid in the exploration of the marker that can lead to early diagnosis. The origin of the increased inflammatory activity in T2DM is virtually unknown, yet the first evidence of a connection between inflammation and diabetes was uncovered more than 100 years ago [9,10].

Biomarkers can be used to help researchers to better grasp the origins of illness. Biological indicators include proteins, genetic and metabolic markers. Inflammatory biomarkers such as orosomucoid, tumour necrosis factor-α (TNF-α), transforming growth factor-B. vascular growth factor endothelial and monocvte chemoattractant protein-1, as well as oxidative stress markers such as 8-hvdroxv-2deoxyguanosine, may be useful for the diagnosis

monitorina of diabetic complications. or Biomarkers can also be employed in biological systems to identify, characterise and observe the expression of proteins. Protein biomarkers are extremely useful to predict long-term mortality in diabetic patients. New biomarkers can be found in tissues and/or biofluids (blood, serum, plasma and urine) [11]. Protein biomarkers have been in biofluids, tissues identified and cells. particularly in T2DM patients [12]. Apolipoproteins, such as apolipoprotein A1, the major component of plasma-bound high-density lipoproteins (HDLs), have been found useful as protein biomarkers [13].

Concentrations of LF in vivo vary according to the type and severity of the disease. Various analytical methods are available to measure these concentrations in order to screen for the presence of diseases such as T2DM, to evaluate their severity, to monitor their progress and to prognoses.Neutrophils offer are polymorphonuclear entities (PMNs) and phagocytic leukocytes that form the first line of defence against invading pathogens in the human body. During inflammation that is induced by tissue injury, they are also essential effector cells [14]. PMNs have roles in chemotaxis, attachment to the endothelium and foreign agents, phagocytosis and microbicidal activity. PMNs have the capacity to enter diseased tissues and destroy invading bacteria by producing a variety of harmful chemicals such as ROS, proteases and LF [15]. The Neutrophils are dysfunctional due to infection agents [16].

The aim of this review is to describe the potential of use of LF as a diagnostic biomarker for T2DM with neutrophil dysfunction and to consider several clinical chemistry analytical techniques that can be used to detect the level of LF in various biological samples. It also highlights the challenges involved.

#### 2. MECHANISMS OF TYPE 2 DIABETES MELLITUS

T2DM is the world's most prevalent and clinically significant metabolic disease. It has become a global epidemic and a huge healthcare burden in recent decades as the number of people with T2DM has increased. In 2013, there were an estimated 382 million T2DM patients worldwide [17], and by 2035, this figure is expected to increase to more than 590 million [18,19].

Diabetes is a "metabolic disease of many etiologies defined by persistent hyperglycemia with disturbances in carbohydrate. lipid, and protein metabolism arising from abnormalities in insulin production, insulin action, or both", according to the World Health Organization [20]. Development of T2DM is closely linked with hereditary variables such as decreased levels of secretion and IR. as well insulin as environmental factors such as obesity, lack of exercise, overeating, stress, inadequate calorie consumption, alcohol use, smoking and ageing [21].

Insulin is produced by β-cells, which first generate pre-proinsulin. During the maturation process, pre-proinsulin undergoes a structural change with the help of many proteins, culminating in production of proinsulin. After that, proinsulin is degraded into C-peptide and insulin. Insulin is retained in granules throughout maturation until insulin release is activated. The release of insulin is largely induced by a reaction to high blood-glucose levels. Other variables, such as levels of amino acids, fatty acids and hormones, can also cause insulin to be released. When blood-glucose levels rise, glucose transporter 2 is used primarily by β-cells to take it in. When glucose enters the cell, it causes the intracellular ratio of adenosine triphosphate to adenosine diphosphate (ATP/ADP) to rise and the ATP-dependent potassium channels in the plasma membrane to close. This process triggers glucose catabolism, which causes the membrane to depolarise and enables  $Ca^{2+}$  to enter the cell through voltage-dependent  $Ca^{2+}$  channels. Insulin exocytosis is triggered by an increase in intracellular  $Ca^{2+}$  concentration, which causes the priming and fusing of secretory insulincontaining granules to the plasma membrane [22,23] (Fig. 1A).

According to recent research, β-cell dysfunction in T2DM may be mediated by a complicated network of interactions between the environment and numerous biochemical processes that occur in the cell. Excessive eating, as with obesity, is associated with hyperglycaemia and hyperlipidaemia, which promote IR and chronic inflammation [24]. The β-cells are subjected to toxic factors such as inflammation, inflammatory stress, metabolic/oxidative stress and amyloid stress under these conditions. These toxic factors can cause loss of islet integrity owing to genetic susceptibility differences [25,26]. Excess amounts of free fatty acids (FFAs) and hyperalycaemia stimulate the apoptotic unfolded protein response pathways, resulting in β-cell malfunction. Obesity-related lipotoxicity. alucotoxicity and glucolipotoxicity induce metabolic and oxidative stress, which lead to βcell death [26,27]. Furthermore, prolonged high levels of glucose enhance proinsulin biosynthesis development of islet and the amyloid polypeptides in  $\beta$ -cells, as well as an increase in production of reactive oxygen species (ROS) [27] (Fig.1B).

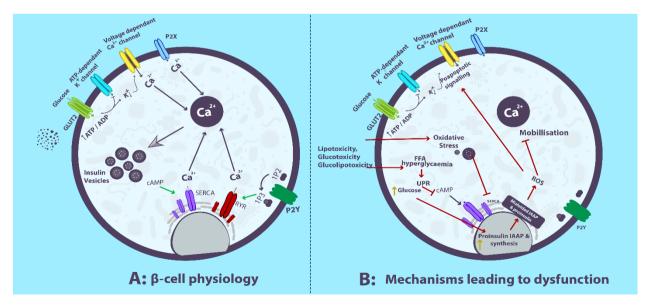


Fig. 1. β-cells in healthy circumstances (A) and during dysfunctional processes (B). Diagram adapted from Galicia-Garcia et al. [28]

# 3. THE ROLE OF INFLAMMATION IN TYPE 2 DIABETES MELLITUS

The first evidence of a connection between inflammation and diabetes was discovered over a century ago. The role of inflammatory processes in the development and progression of T2DM has received greater attention than it has in type 1 diabetes due to an important mechanism. Increases in levels of inflammatory markers have been seen in seemingly healthy people who subsequently acquire T2DM. This finding suggests that inflammation starts early in the period of reduced glucose tolerance and prior to T2DM diagnosis [24,29,30].

IR has long been thought to be a key factor in the pathophysiology and progression of T2DM. IR begins prior to the onset of T2DM, when  $\beta$ -cell breakdown and insulin insufficiency result in decreased glucose tolerance. Several elements, including genetics and environmental effects, obesity, and other diseases associated with chronic inflammation or infection, have been related to the development of IR in people with impaired glucose tolerance and T2DM [24].

Fig. 2 shows how inflammation develops in T2DM, as described by Donath and Shoelson [29]. Overeating causes levels of blood glucose and FFAs to rise, and this leads to metabolic stress in various tissues. This stress triggers the production of a variety of pro-inflammatory

cytokines and chemokines. Immune cells are drawn in, and this process contributes to tissue inflammation [24,29,30].

IR is linked primarily to a variety of proinflammatory and/or oxidative stress mediators, including pro-inflammatory cytokines such as interleukins (ILs) 1 $\beta$  and 6, and TNF- $\alpha$ , as well as a variety of chemokines and adipocytokines [31,32]. These pro-inflammatory cytokines can cause systemic insulin sensitivity and glucose imbalance as they directly increase IR in adipocytes, muscle cells and hepatic cells. Increased levels of these pro-inflammatory cytokines cause the liver to generate and release plasminogen C-reactive protein, activator inhibitor-1, amyloid-A, 1-acid glycoprotein and haptoglobin. These proteins first appear in the early stages of T2DM, and their blood levels grow as the disease progresses [33].

Several studies have revealed the occurrence of various inflammatory responses in  $\beta$ -cells and peripheral tissues. These studies report that IL- $\beta$  is a master pro-inflammatory mediator that activates a plethora of other pro-inflammatory cytokines and chemokines. Once inflammation is triggered, it has a negative impact on  $\beta$ -cells in pancreatic islets, resulting in decreased insulin production. Similarly, IL- $\beta$  plays a critical role in the induction of inflammation in peripheral tissues, which contributes to the development of IR in these tissues [34,35].

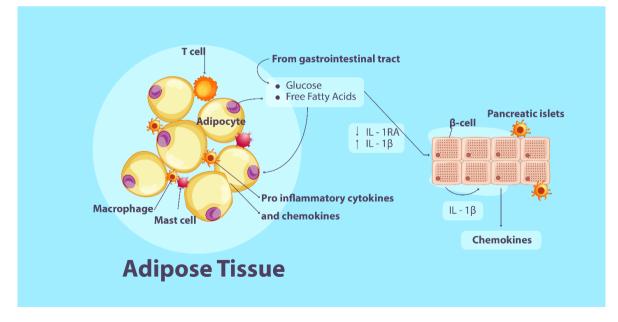


Fig. 2. How inflammation develops in T2DM. Diagram adapted from Donath et al. [29]

TNF- $\alpha$  has been identified as a key cytokine that is implicated in IR. TNF-a has metabolic effects in peripheral tissue as it alters the expression of genes involved in lipolysis and lipogenesis, which results in a rise in FFA concentrations. Higher levels of hepatic gluconeogenesis and IR in skeletal muscle are linked with increased FFA levels. This increase in FFAs is also linked with insulin hypersecretion, which can lead to a reduction in insulin secretion capacity [36]. TNF- $\alpha$  plays an essential role in insulin signalling system as it inhibits activity of the insulin receptor tyrosine kinase in adipocytes, which leads to decreased phosphorylation and activation of insulin receptor substrate-1 and so limits cell responsiveness to insulin. TNF-α has also been linked to a reduction in the expression of genes that encode proteins that produce insulin receptor substrates [37].

Ageing is linked to higher amounts of IL-6 in the blood, which can be linked with increased IR [38]. The process by which IL-6 triggers IR is complex and multifaceted. It not only stops nonoxidative glucose from being metabolised, but it also inhibits the activity of lipoprotein lipase, which raises triglyceride levels in the blood [39]. Furthermore, the presence of IL-6 activates the cytokine signalling suppressor, which may inhibit the activation of cytokine-mediated insulin receptor transcriptional factors [40]. As a result, IL-6 is considered a critical biomarker for IR development.

# 4. NEUTROPHILS AS BIOMARKERS IN DM

Interactions with the vascular endothelium regulate neutrophil migration from the circulation to the site of inflammation. Primed neutrophils actively manufacture and release cytokines. chemokines, leucotrienes and prostaglandins, and present antigens, by virtue of their vast numbers inside inflammatory tissue, which leads to local generation of inflammatory mediators. In response to a variety of stimuli, including TNF- $\alpha$ , neutrophils have been demonstrated to manufacture and release IL-8 [41,42]. Activated neutrophils have also been shown to produce IL-1, IL-6, IL-12, TNF- $\alpha$  and oncostatin M, all of which can stimulate the activity of neutrophils and other immune-system cells [43]. It is important to find out where various neutrophil phenotypes are made durina severe inflammation events. Neutrophil cell-surface markers or their derivatives can be employed as biomarkers in disease diagnosis. Chronic inflammation is a feature of T2DM, which involves humoral components as well as several kinds of white blood cells, such as mononuclear and PMN leukocytes. The development of T2DM has been linked to an increase in neutrophil count and phagocytic dysfunction. This is related to the well-known theory that oxidative stress, which is generally created by neutrophil activity, causes diabetes problems [44].

T2DM is now recognised as an inflammatory condition that is associated with IR and abnormal endothelial vascular reactivity. Insulin has been shown to have a substantial regulatory influence on the functional activities of immune cells [45,46]. Insulin's priming effect on PMN activity can be viewed as the body creating a wide defence to support the major immunological response to antigen exposure, and this response is aided by meal intake [47]. Insulin sensitivity declines with age, and this situation adds to the immune system's age-related deterioration, particularly after meal consumption [46].

PMN function is influenced by the conditions caused by T2DM, age-related IR, diet and lifestyle. In human and animal models of diabetes, abnormalities in neutrophil adhesion, chemotaxis, phagocytosis, ROS generation and microbicidal activity have been reported [48,49]. The occurrence of hyperglycaemia reduces the activity of glucose-6-phosphate dehydrogenase (G6PD) and glutaminase enzymes, while it increases the activity of phosphofructokinase [48]. Reduced G6PD activity impairs the development of the pentose-phosphate pathway as well as neutrophil activities [50]. Even when the subject's glycaemic index is incorrect, insulin enhances neutrophil phagocytosis and ROS generation. This finding suggests that insulin has a direct effect on neutrophils [48]. Furthermore, increased levels of circulating FFA and triacylglycerol promote IR as well as neutrophilic inflammation [51,52].

Changes in immune-cell activity may explain why the T2DM and older populations suffer infections more frequently than other people. Research has demonstrated that treatment of hyperglycaemia with insulin can lead to restoration of diabetic patients' impaired PMN functioning. Glucose intake and glycogen metabolism in PMNs are both insulin-dependent, despite the fact that PMNs do not require insulin in order to absorb glucose. Following insulin therapy, insulin receptor expression has been found to be linked to PMN chemotaxis in both young and old individuals. In insulin-resistant and elderly people, antimicrobial protein synthesis in PMNs is altered, and it is reduced in all humans after endotoxin iniections intravenous under hyperglycaemic conditions [15]. Elgazar-Carmon et al. (2008) discovered that a high-fat diet caused significant neutrophil recruitment to intraabdominal adipocyte tissue; this recruitment peaked after three to seven days and subsequently faded. The researchers theorised that neutrophil recruitment was necessary to kickstart the inflammatory response to high-fat meals. These neutrophils may produce chemotactic factors, which enable macrophage infiltration and the continuation of an inflammatory state in adipose tissue. This chronic inflammatory infiltration is preceded by a short, acute infiltration of inflammatory molecules that are dominated by neutrophils, according to a well-established paradigm in systemic inflammatory processes [53].

#### 5. NEUTROPHIL DYSFUNCTION IN TYPE 2 DIABETES: SPECIFIC MARKERS

In patients with IR or T2DM, levels of antibacterial neutrophil proteins such as LF, bactericidal/increasing permeability protein (BPI) and  $\alpha$ -defensins are decreased. The decreased antibacterial ability of neutrophils that occurs in T2DM patients has been found to correspond with the circulating levels of these proteins [54].

BPI is a 55kDa cationic protein that is found in the azurophilic granules of neutrophils. Plasma BPI concentration has been found to be negatively related to metabolic indices and directly correlated with insulin sensitivity and levels of HDLs [55,56].

Human  $\alpha$ -defensins are peptides that contain 29-35 amino acids and have high arginine content. In seemingly healthy Caucasian males, significant positive correlations have been found between concentrations of plasma  $\alpha$ -defensin, insulin sensitivity, nonatherogenic lipid profile and vascular function [57,58].

Furthermore, numerous investigations have shown that development of T2DM is linked to a change in neutrophil functioning (lower bactericidal ability and higher neutrophil count) [59-61].

#### 6. LACTOFERRIN AS A DIAGNOSTIC MARKER FOR TYPE 2 DIABETES DISEASES

LF levels in the body are elevated during development of an infection or an inflammatory

disease, which means that LF can be used as a biomarker for inflammatory disorders. The presence of LF also reduces inflammation as it interacts with macrophages and limits the production of inflammatory cytokines by cells in a similar way to other anti-inflammatory cytokines, according to several studies [62]. Scientists have made many attempts to improve their comprehension of the role of LF in the maintenance of human health [63].

Videm et al. (2007) reported that when a person who displayed traditional risk factors for coronary artery disease was infected with Chlamydia. pneumoniae, coronary artery disease would develop only if monocytes/macrophages and neutrophils were activated. According to this study, increased concentrations of LF but not of myeloperoxidase are linked with the occurrence of severe coronary artery stenosis [64]. Furthermore, both at baseline and after a fat overload, circulating concentrations of LF have been reported to be negatively correlated in extremely obese individuals with postprandial lipaemia and production of oxidative stress glutathione markers catalase and (e.g., peroxidase) and C-reactive protein [65].

According to recent studies, LF can be utilised as a biomarker in the detection of inflammatory bowel disease (IBD) [66], Alzheimer's disease (AD) [67] and dry-eye disease (DED) [68]. Clinical grading systems and endoscopy have traditionally been used to diagnose IBD. However, both these methods are costly and show limited accuracy. Previous research has suggested that faecal levels of LF might be useful biomarkers to predict the development of IBDs [69]. As PMN neutrophils degranulate intestinal inflammation. durina secondarv granules are produced. LF is a key component of secondary granules. and therefore LF concentration increases in cases of IBD. In cases of Crohn's disease (CD) and ulcerative colitis (UC) in children, the levels of faecal LF have been reported to be greater than those in control participants (7.3g/g), although the diagnostic efficacy of this protein in UC patients is reported to be better than in CD patients [70].

It is difficult to diagnose AD early in its development. Current tactics involve combining the techniques of positron emission tomography and magnetic resonance imaging to assess the levels of tau and amyloid proteins in the cerebrospinal fluid [71]. There have been attempts to create a rapid and cost-effective diagnostic method. Research in AD pathophysiology suggests that bacterial and viral infections may induce onset of the disease and lead to a weakened innate immune system [72]. Saliva is considered the body's first line of defence against infection because it contains numerous antimicrobial proteins. Some reports have linked oral infections to the development of AD [73]. LF is an essential defence component of saliva due to its particular antibacterial properties. Therefore, the measurement of salivary LF levels may offer a diagnostic method for the early detection of AD. González- Sánchez et al. (2020) measured levels of salivary LF in order to diagnose prodromal AD and to study the relationship between salivary LF levels and cerebral amyloid-B. The results showed that salivary LF levels did not decrease in other dementias, such as frontotemporal dementia, and that reduced levels of LF could be attributed to the disruption of hypothalamic function due to early hypothalamic amyloid- $\beta$  accumulation [67].

DED, a common ocular surface disease of multifactorial aetiology, causes many symptoms and visual impairment, sometimes with ocular surface damage [74]. DED is currently diagnosed through use of several tests such as evaluation of the tear osmolarity, the Schirmer tear test and the phenol red thread test [75]. However, these methods are of low accuracy and can be easily affected by environmental factors. The presence of LF in the tear film plays a key role in the avoidance of ocular diseases because of its unique antimicrobial and anti-inflammatory activities [76]. Some recent research has confirmed that the concentrations of LF in tears are significantly different between patients with DED and controls [67,77].

A 1995 study discovered that mesencephalon samples that had been obtained by autopsy from eight patients with histologically confirmed Parkinson's disease (PD) showed a higher content of the LF receptor than samples taken from 13 people with no known history of psychiatric or neurological disorders. This finding kicked off research into a link between levels of LF and the pathogenesis of PD [78]. Additional examination of the mesencephalon cellular distribution revealed significant levels of LF in a wide population of neurons in the substantia nigra (SN) of control individuals. In comparison with control instances, individuals with PD exhibited greater LF levels in the surviving neurons of the SN, according to quantitative analyses. The researchers concluded: "Further

research will be required to understand whether LF serves as an iron scavenger and may represent a protective factor, or whether it promotes excessive iron buildup leading to oxidative injury in susceptible neurons" [79]. Two recent investigations separately showed that LF might be useful as a non-invasive PD marker, after they discovered that the levels of LF in the saliva and tears of PD patients were higher than those in the same biofluids of the control group [80,81]. Salivary and lacrimal LF levels could be acceptable as PD markers since they are simpler to collect than blood samples and, more importantly, the amounts of LF in both exocrine secretions are substantially larger than the levels of oligomeric alpha-synuclein. This compound is widely utilised as a marker despite its high prevalence in red blood cells, low concentration in biological fluids and contradictory metaanalysis results, which limit its utility in PD diagnosis [82].

Several studies have shown that LF regulates the production of inflammatory cytokines in a similar way to other anti-inflammatory cytokines. LF has been found to reduce  $\beta$ -cell damage by decreasing production of TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and IL-8 in human mononuclear cells (*in vitro*) while it increases IL-10 and IL-4 production (*in vivo*) [83].

#### 7. CORRELATION OF LF ANALYSIS WITH INSULIN RESISTANCE AND TYPE 2 DIABETES STUDIES

The observation that there was a negative connection between concentrations of circulating LF and of fasting glucose [84,85] and that there was a positive correlation between levels of circulating LF and insulin sensitivity [85] sparked research into the role of LF in glucose metabolism changes. It is possible that LF has a direct impact on IR in peripheral organs [86]. Mohamed et al. (2018) discovered that glucose metabolism in diabetic children was changed compared with that in their control counterparts and that the diabetic youngsters showed a twofold increase in LF levels [87]. The increased levels of LF helped weight loss by improving insulin action and increasing the activity of the fuel-sensing protein [65].

## 8. NEUTROPHIL DYSFUNCTION AND LF PRODUCTION

In a broad sense, LF is an acute-phase protein that functions as an "alarmin". Alarmins form a

#### Table 1. Selected studies of quantitative measurements of LF in T2DM patients

Study	Sample type	Analytical method	Conclusions	Reference
LF concentrations in tears and tear secretion rates were measured in normal and diabetic individuals.	Tears	ELISA	There was no link between the amount of LF in the tears of the diabetic subjects and the length of time they had been diabetic, and there was no variation in the amount of LF in normal tears vs. diabetic tears.	[97]
Diabetic and diabetic with retinopathy patients randomly selected for study of tears.	Tears	SDS-PAGE	Tear film was decreased more in diabetic subjects than in normal subjects.	[98]
In hamsters in which diabetes was induced with streptozotocin, the levels of salivary antibacterial agents such as lysozyme, lactoperoxidase and LF were measured.	Saliva	Gel electro- phoresis	Ratio of LF to total protein in the hamsters was about twice that of the control hamsters. Insulin therapy restored 73% and 74% of the activity of lysozyme and lactoperoxidase, respectively, and the ratio of LF to total salivary protein returned to normal levels.	[99]
The connection between circulating LF, LF gene polymorphisms, dyslipidaemia and vascular reactivity in male humans with glucose intolerance was examined.	Plasma	ELISA	With IR and T2DM, the concentration of circulating LF was reduced. Fasting triglyceride concentration, body-mass index, waist-to-hip ratio, and fasting glucose levels were all shown to be inversely associated with levels of LF, whereas HDL cholesterol concentration was found to be directly related.	[84]
In a Caucasian population, the association between circulating LF and chronic inflammation-associated IR was investigated according to glucose tolerance level.	Plasma	ELISA	LF levels in the blood were found to be substantially lower in patients with impaired glucose tolerance compared with healthy people. It was possible that LF was involved in persistent low-level inflammation and IR.	[85]
The goal of this study was to see how well levels of myeloperoxidase and LF, two neutrophil degranulation products, predicted long-term risk of fatal ischaemic heart disease in patients with T2DM and in healthy people.	Serum	ELISA	There was no significant difference in LF levels between T2DM patients and controls.	[100]
Concentrations of LF were measured in T2DM patients compared with non-diabetic controls.	Serum	ELISA	Levels of LF were greater In T2DM patients than in control participants.	[101]
The concentrations were measured of protective factors in the saliva of individuals with T2DM who had decompensated.	Saliva	ELISA	In decompensated T2DM patients, salivary LF levels were significantly lower than in the control group.	[102]
LF was studied as a biochemical marker in individuals with T2DM and in those with T2DM and peripheral neuropathy (diabetic nerve pain, DNP).	Serum	ELISA	T2DM patients showed significantly higher serum LF levels when compared with the control group, whereas those with DNP showed highly significant increases when compared with both the control and T2DM groups. LF was likewise favourably associated with levels of HbA1c in the T2DM group and negatively with Fe in the DNP group.	[103]

small family of proteins that are produced by neutrophils in response to infection. They play a key role in the modification of immune reactivity in response to a pathogenic encounter or clinical insult [88,89].

LF is expressed and stored in secondary granules by neutrophil leukocytes, which make up more than half of all white blood cells. LF is produced in the blood during neutrophil activation, which occurs at the initial stages of attachment to the activated endothelium. Its concentration can reach 200mg/l (compared with around 1mg/l under normal conditions). especially in inflamed tissues. Microglial cells, which function as resident macrophages in the brain, also produce LF when the brain is inflamed [90]. In a confined area, LF as an alarmin develops conditional connections between neutrophils and dendritic cells [88].

The innate system's armoury that is used to establish microbial balance in mucosal fluids includes LF, released immunoglobulin A and defensins [91]. LF is a multifunctional molecule due to its tendency to interact with microbial and target host-cell surfaces and its high affinity for ferric iron. This affinity deprives bacteria of the free iron they require to flourish [92]. The antibacterial properties of neutrophilic LF, which is generated in high concentration in infected tissues and is probably linked to the chromatin fibril matrix released by neutrophils (neutrophil extracellular traps), are comparable [93].

#### 9. METHODS OF BIOANALYSIS OF LF IN DIABETES PATIENTS

Various bioanalytical techniques have been used to measure levels of LF in biological samples. One of the most popular, due to its specificity and sensitivity, is enzyme-linked immunosorbent assay (ELISA). It is a quantitative, highly accurate, fast technique that can detect molecules in ng/ml concentrations. ELISA can be used to detect the binding of analyte and specific antibodies [94]. ELISA techniques have been reported in many studies of LF levels in DED, Crohn's disease and diabetes [95-97]. Table 1 highlights the evidence that is shown in selected studies and which supports the existence of a relationship between levels of indicators of neutrophil dysfunction (LF) and T2DM. The table also includes references to LF testina techniques. According to the findings shown in the table, T2DM is associated with decreased LF production and/or secretion in neutrophils.

#### **10. CONCLUSION**

LF is a protein that plays an important role in inflammation. In many studies, it has been found to play a key role in the development of T2DM. Due to neutrophil dysfunction, LF levels usually affect clinical diagnosis. ELISA techniques have been used widely to detect the concentrations of LF in diabetic patients due to their high sensitivity and selectivity. However, use of other techniques such as LC-Ms/Ms and proteomics should be considered to improve the analysis and understanding of LF activity and quantity in diabetes.

#### DISCLAIMER

Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

#### CONSENT AND ETHICAL APPROVAL

It is not applicable.

#### **COMPETING INTERESTS**

Author has declared that no competing interests exist.

#### REFERENCES

- 1. Johanson B. Isolation of an iron-containing red protein from human milk. Acta Chemica Scandinavica. 1960;14:510-512.
- 2. Legrand D, Elass E, Carpentier M, Mazurier J. Lactoferrin. Cellular and Molecular Life Sciences. 2005;62:2549-2559.
- 3. Mayeur S, Veilleux A, Pouliot Y, Lamarche B, Beaulieu J-F, Hould FS, Richard D, Tchernof A, Levy E. Plasma Lactoferrin levels positively correlate with insulin resistance despite an inverse association with total adiposity in lean and severely obese patients. PloS One. 2016;11: e0166138-e0166138,

DOI: 10.1371/journal.pone.0166138

- González-Chávez, SA, Arévalo-Gallegos S, Rascón-Cruz Q. Lactoferrin: Structure, function and applications. International Journal of Antimicrobial Agents. 2009;33: 301-e301.
- 5. Kruzel ML, Zimecki M, Actor JK. Lactoferrin in a context of inflammation-

induced pathology. Frontiers in Immunology. 2017;8:1438.

- Shepard BD. Sex differences in diabetes and kidney disease: Mechanisms and consequences. American Journal of Physiology-Renal Physiology. 2019;317: F456-F462.
- 7. Force IDFCGT. Global guideline for type 2 diabetes. Brussels: International Diabetes Federation; 2005.
- Waugh N, Scotland G, McNamee P, Gillett M, Brennan A, Goyder E, Williams R, John A. Screening for type 2 diabetes: Literature review and economic modelling. Health Technology Assessment-Southampton-2007, 11.
- 9. Reid J, Macdougall AI, Andrews MM. Aspirin and diabetes mellitus. British Medical Journal. 1957; 2:1071.
- 10. Williamson RT. On the treatment of glycosuria and diabetes mellitus with sodium salicylate. British Medical Journal. 1901;1:760.
- 11. Ndisang JF, Rastogi S, Vannacci A. Insulin resistance, type 1 and type 2 diabetes, and related complications 2015; 2015.
- Riaz S, Alam SS, Srai SK, Skinner V, Riaz A, Akhtar MW. Proteomic identification of human urinary biomarkers in diabetes mellitus type 2. Diabetes Technology & Therapeutics. 2010;12:979-988.
- Davidsson P, Hulthe J, Fagerberg Br, Camejo G. Proteomics of apolipoproteins and associated proteins from plasma highdensity lipoproteins. Arteriosclerosis, Thrombosis, and Vascular Biology. 2010; 30:156-163.
- 14. Nothan C. Neutrophils and immunity: Challenges and port unities. Nat. Rev. Immunol. 2006;6:173-173.
- Walrand S, Guillet C, Boirie Y, Vasson M-P. Insulin differentially regulates monocyte and polymorphonuclear neutrophil functions in healthy young and elderly humans. The Journal of Clinical Endocrinology & Metabolism. 2006;91: 2738-2748.
- Dinauer MC. Disorders of neutrophil function: An overview. Methods in molecular biology (Clifton, N.J.). 2014; 1124:501-515. DOI: 10.1007/978-1-62703-845-4 30
- 17. Reed J, Bain S, Kanamarlapudi V. A review of current trends with type 2 diabetes epidemiology, aetiology, pathogenesis, treatments and future perspectives. Diabetes, Metabolic

Syndrome and Obesity: Targets and Therapy. 2021;14:3567-3567.

- Boyle JP, Thompson TJ, Gregg EW, Barker LE, Williamson DF. Projection of the year 2050 burden of diabetes in the US adult population: Dynamic modeling of incidence, mortality, and prediabetes prevalence. Population Health Metrics. 2010;8:1-12.
- 19. Ozougwu JC, Obimba KC, Belonwu CD, Unakalamba CB. The pathogenesis and pathophysiology of type 1 and type 2 diabetes mellitus. Journal of Physiology and Pathophysiology. 2013;4:46-57.
- 20. Thompson A, Kanamarlapudi V. Type 2 diabetes mellitus and glucagon like peptide-1 receptor signalling. Clin Exp Pharmacol. 2013;3:1459-2161.
- 21. Kohei K. Pathophysiology of type 2 diabetes and its treatment policy. JMAJ. 2010;53:41-46.
- 22. Fu Z; R Gilbert E, Liu D. Regulation of insulin synthesis and secretion and pancreatic Beta-cell dysfunction in diabetes. Current Diabetes Reviews. 2013; 9:25-53.
- Rorsman P, Ashcroft FM. Pancreatic β-cell electrical activity and insulin secretion: of mice and men. Physiological Reviews. 2018;98:117-214.
- 24. Czech MP. Insulin action and resistance in obesity and type 2 diabetes. Nature Medicine. 2017;23:804-814.
- 25. Christensen AA, Gannon M. The beta cell in type 2 diabetes. Current Diabetes Reports. 2019;19:1-8.
- Halban PA, Polonsky KS, Bowden DW, Hawkins MA, Ling C, Mather KJ, Powers AC, Rhodes CJ, Sussel L, Weir GC. β-cell failure in type 2 diabetes: Postulated mechanisms and prospects for prevention and treatment. The Journal of Clinical Endocrinology & Metabolism. 2014;99: 1983-1992.
- Yamamoto WR, Bone RN, Sohn P, Syed F, Reissaus CA, Mosley AL, Wijeratne AB, True JD, Tong X, Kono T. Endoplasmic reticulum stress alters ryanodine receptor function in the murine pancreatic β cell. Journal of Biological Chemistry. 2019;294: 168-181.
- Galicia-Garcia U, Benito-Vicente A, Jebari S, Larrea-Sebal A, Siddiqi H, Uribe KB, Ostolaza H, Martín C. Pathophysiology of type 2 diabetes mellitus. International Journal of Molecular Sciences. 2020;21: 6275-6275.

DOI: 10.3390/ijms21176275

- 29. Donath MY, Shoelson SE. Type 2 diabetes as an inflammatory disease. Nature Reviews Immunology. 2011;11:98-107.
- Hu FB, Meigs JB, Li TY, Rifai N, Manson JE. Inflammatory markers and risk of developing type 2 diabetes in women. Diabetes. 2004;53:693-700.
- 31. Rehman K, Akash MSH. Mechanisms of inflammatory responses and development of insulin resistance: How are they interlinked? Journal of Biomedical Science. 2016;23:1-18.
- 32. King GL. The role of inflammatory cytokines in diabetes and its complications. Journal of Periodontology. 2008;79:1527-1534.
- Festa A, D'Agostino R, Tracy RP, Haffner SM. Elevated levels of acute-phase proteins and plasminogen activator inhibitor-1 predict the development of type 2 diabetes: The insulin resistance atherosclerosis study. Diabetes. 2002;51: 1131-1137.
- Akash MSH, Shen Q, Rehman K, Chen S. Interleukin-1 receptor antagonist: A new therapy for type 2 diabetes mellitus. Journal of Pharmaceutical Sciences. 2012; 101:1647-1658.
- Jager J, Grémeaux T, Cormont M, Le Marchand-Brustel Y, Tanti J-F. Interleukin-1β-induced insulin resistance in adipocytes through down-regulation of insulin receptor substrate-1 expression. Endocrinology. 2007;148:241-251.
- Swaroop JJ, Rajarajeswari D, Naidu JN. Association of TNF-α with insulin resistance in type 2 diabetes mellitus. The Indian Journal of Medical Research. 2012; 135:127-127.
- Davis R, Aguirre V, Uchida T, Yenush L, White MF. The c-Jun NH2-terminal kinase promotes insulin resistance during association with insulin receptor substrate-1 and phosphorylation of Ser307. Journal of Biological Chemistry. 2000;275:9047-9054.
- Abbatecola AM, Ferrucci L, Grella R, Bandinelli S, Bonafè M, Barbieri M, Corsi AM, Lauretani F, Franceschi C, Paolisso G. Diverse effect of inflammatory markers on insulin resistance and insulin-resistance syndrome in the elderly. Journal of the American Geriatrics Society. 2004;52:399-404.
- 39. Kern PA, Ranganathan S, Li C, Wood L, Ranganathan G. Adipose tissue tumor

necrosis factor and interleukin-6 expression in human obesity and insulin resistance. American Journal of Physiology-Endocrinology and Metabolism. 2001;280:E745-E751.

- 40. Tilg H, Moschen AR. Inflammatory mechanisms in the regulation of insulin resistance. Molecular Medicine. 2008;14: 222-231.
- 41. Wright HL, Moots RJ, Bucknall RC, Edwards SW. Neutrophil function in inflammation and inflammatory diseases. Rheumatology. 2010;49:1618-1631.
- 42. Wright HL, Moots RJ, Edwards SW. The multifactorial role of neutrophils in rheumatoid arthritis. Nature Reviews Rheumatology. 2014;10:593-601.
- 43. Cross A, Edwards SW, Bucknall RC, Moots RJ. Secretion of oncostatin M by neutrophils in rheumatoid arthritis. Arthritis & Rheumatism. 2004;50:1430-1436.
- 44. Carlson M, Raab Y, Seveus L, Xu S, Hällgren R, Venge P. Human neutrophil lipocalin is a unique marker of neutrophil inflammation in ulcerative colitis and proctitis. Gut. 2002;50:501-506.
- 45. Okouchi M, Okayama N, Shimizu M, Omi H, Fukutomi T, Itoh M. High insulin exacerbates neutrophil-endothelial cell adhesion through endothelial surface expression of intercellular adhesion molecule-1 via activation of protein kinase C and mitogen-activated protein kinase. Diabetologia. 2002;45:556-559.
- 46. Walrand Š, Guillet C, Boirie Y, Vasson MP. In vivo evidences that insulin regulates human polymorphonuclear neutrophil functions. Journal of Leukocyte Biology. 2004;76:1104-1110.
- 47. Walrand S, Moreau K, Caldefie F, Tridon A, Chassagne J, Portefaix G, Cynober L, Beaufrère B, Vasson M-P, Boirie Y. Specific and nonspecific immune responses to fasting and refeeding differ in healthy young adult and elderly persons. The American Journal of Clinical Nutrition. 2001;74:670-678.
- 48. Alba-Loureiro TC, Munhoz CD, Martins JO, Cerchiaro GA, Scavone C, Curi R, Sannomiya P. Neutrophil function and metabolism in individuals with diabetes mellitus. Brazilian Journal of Medical and Biological Research. 2007;40:1037-1044.
- 49. Alba-Loureiro TC, Hirabara SM, Mendonca JR, Curi R, Pithon-Curi TC. Diabetes causes marked changes in function and

metabolism of rat neutrophils. Journal of Endocrinology. 2006;188:295-303.

- 50. Perner A, Nielsen SE, Rask-Madsen J. High glucose impairs superoxide production from isolated blood neutrophils. Intensive Care Medicine. 2003;29:642-645.
- Sina C, Gavrilova O, Förster M, Till A, Derer S, Hildebrand F, Raabe B, Chalaris A, Scheller J, Rehmann A. G proteincoupled receptor 43 is essential for neutrophil recruitment during intestinal inflammation. The Journal of Immunology. 2009;183:7514-7522.
- 52. Vinolo MAR, Ferguson GJ, Kulkarni S, Damoulakis G, Anderson K, Bohlooly-Y M, Stephens L, Hawkins PT, Curi R. SCFAs induce mouse neutrophil chemotaxis through the GPR43 receptor. PloS One. 2011;6:e21205-e21205.
- 53. Elgazar-Carmon V, Rudich A, Hadad N, Levy R. Neutrophils transiently infiltrate intra-abdominal fat early in the course of high-fat feeding. Journal of Lipid Research. 2008;49:1894-1903.
- 54. Moreno-Navarrete JM, Fernández-Real JM. Antimicrobial-sensing proteins in obesity and type 2 diabetes: The buffering efficiency hypothesis. Diabetes Care. 2011;34(Suppl 2):S335-S341. DOI: 10.2337/dc11-s238
- 55. Wittmann I, Schönefeld M, Aichele D, Groer G, Gessner A, Schnare M. Murine bactericidal/permeability-increasing protein inhibits the endotoxic activity of lipopolysaccharide and gram-negative bacteria. The Journal of Immunology. 2008;180:7546-7552.
- 56. Sun Q, Li T, Li Y, Wei L, Zhang M, Deng S. Bactericidal/permeabilityincreasing protein improves cognitive impairment in diabetic mice via blockade of the LPS-LBP-TLR4 signaling pathway. Frontiers in Physiology. 2021;11:718-718.
- López-Bermejo A, Chico-Julia B, Castro A, Recasens M, Esteve E, Biarnés J, Casamitjana R, Ricart W, Fernández-Real J-M. Alpha defensins 1, 2, and 3: Potential roles in dyslipidemia and vascular dysfunction in humans. Arteriosclerosis, Thrombosis, and Vascular Biology. 2007; 27:1166-1171.
- Schneider JJ, Unholzer A, Schaller M, Schäfer-Korting M, Korting HC. Human defensins. Journal of Molecular Medicine. 2005;83:587-595.
- 59. Advani A, Marshall S, Thomas T. Impaired neutrophil actin assembly causes

persistent CD11b expression and reduced primary granule exocytosis in Type II diabetes. Diabetologia. 2002;45:719-727.

- 60. Shim WS, Kim HJ, Kang ES, Ahn CW, Lim SK, Lee HC, Cha BS. The association of total and differential white blood cell count with metabolic syndrome in type 2 diabetic patients. Diabetes Research and Clinical Practice. 2006;73:284-291.
- 61. Shurtz-Swirski R, Sela S, Herskovits AT, Shasha SM, Shapiro G, Nasser L, Kristal B. Involvement of peripheral polymorphonuclear leukocytes in oxidative stress and inflammation in type 2 diabetic patients. Diabetes Care. 2001;24:104-110.
- 62. Yamaguchi M, Matsuura M, Kobayashi K, Sasaki H, Yajima T, Kuwata T. Lactoferrin protects against development of hepatitis caused by sensitization of Kupffer cells by lipopolysaccharide. Clinical Diagnostic Laboratory Immunology. 2001;8:1234-1239.
- Agrawal RP, Dogra R, Mohta N, Tiwari R, Singhal S, Sultania S. Beneficial effect of camel milk in diabetic nephropathy. Acta Biomedica de l'Ateneo Parmense. 2009; 80:131-134.
- 64. Videm V, Wiseth R, Gunnes S, Madsen HO, Garred P. Multiple inflammatory markers in patients with significant coronary artery disease. International Journal of Cardiology. 2007;118:81-87.
- 65. Fernández-Real JM, García-Fuentes E, Moreno-Navarrete JM, Murri-Pierri M, Garrido-Sánchez L, Ricart W, Tinahones F. Fat overload induces changes in circulating lactoferrin that are associated with postprandial lipemia and oxidative stress in severely obese subjects. Obesity. 2010;18:482-488.
- Kane SV, Sandborn WJ, Rufo PA, Zholudev A, Boone J, Lyerly D, Camilleri M, Hanauer SB. Fecal lactoferrin is a sensitive and specific marker in identifying intestinal inflammation. The American Journal of Gastroenterology. 2003;98: 1309-1314.
- González-Sánchez M, Bartolome F, Antequera D, Puertas-Martín V, González P, Gómez-Grande A, Llamas-Velasco S, Herrero-San Martín A, Pérez-Martínez D, Villarejo-Galende A. Decreased salivary lactoferrin levels are specific to Alzheimer's disease. EBioMedicine. 2020;57:102834-102834.
- 68. Narayanan S, Redfern RL, Miller WL, Nichols KK, McDermott AM. Dry eye

disease and microbial keratitis: Is there a connection? The Ocular Surface. 2013;11: 75-92.

- Angriman I, Scarpa M, D'Incà R, Basso D, Ruffolo C, Polese L, Sturniolo GC, D'Amico DF, Plebani M. Enzymes in feces: Useful markers of chronic inflammatory bowel disease. Clinica Chimica Acta. 2007;381: 63-68.
- Buderus S, Boone JH, Lentze MJ. Fecal lactoferrin: Reliable biomarker for intestinal inflammation in pediatric IBD. Gastroenterology Research and Practice. 2015;2015.
- Simonsen AH, Herukka S-K, Andreasen N, Baldeiras I, Bjerke M, Blennow K, Engelborghs S, Frisoni GB, Gabryelewicz T, Galluzzi S. Recommendations for CSF AD biomarkers in the diagnostic evaluation of dementia. Alzheimer's & Dementia. 2017;13:274-284.
- 72. Sun X-W, Liu C-M, Teng Z-Q. Commentary: Multiscale analysis of independent alzheimer's cohorts finds disruption of molecular, genetic, and clinical networks by human herpesvirus. Frontiers in Molecular Neuroscience. 2018; 11:340-340.
- Welling MM, Nabuurs RJA, van der Weerd L. Potential role of antimicrobial peptides in the early onset of Alzheimer's disease. Alzheimer's & Dementia. 2015;11:51-57.
- 74. Craig JP, Nichols KK, Akpek EK, Caffery B, Dua HS, Joo C-K, Liu Z, Nelson JD, Nichols JJ, Tsubota K. TFOS DEWS II definition and classification report. The Ocular Surface. 2017;15:276-283.
- Lemp MA, Bron AJ, Baudouin C, Del Castillo JMB, Geffen D, Tauber J, Foulks GN, Pepose JS, Sullivan BD. Tear osmolarity in the diagnosis and management of dry eye disease. American Journal of Ophthalmology. 2011;151:792-798.
- Flanagan JL, Willcox MDP. Role of lactoferrin in the tear film. Biochimie. 2009; 91:35-43.
- 77. Versura P, Nanni P, Bavelloni A, Blalock WL, Piazzi M, Roda A, Campos EC. Tear proteomics in evaporative dry eye disease. Eye. 2010;24:1396-1402.
- 78. Faucheux BA, Nillesse N, Damier P, Spik G, Mouatt-Prigent A, Pierce A, Leveugle B, Kubis N, Hauw J-J, Agid Y. Expression of lactoferrin receptors is increased in the mesencephalon of patients with Parkinson

disease. Proceedings of the National Academy of Sciences. 1995;92:9603-9607.

- 79. Leveugle B, Faucheux BA, Bouras C, Nillesse N, Spik G, Hirsch EC, Agid Y, Hof PR. Cellular distribution of the ironbinding protein lactotransferrin in the mesencephalon of Parkinson's disease cases. Acta Neuropathologica. 1996;91: 566-572.
- Hamm-Alvarez SF, Janga SR, Edman MC, Feigenbaum D, Freire D, Mack WJ, Okamoto CT, Lew MF. Levels of oligomeric α-Synuclein in reflex tears distinguish Parkinson's disease patients from healthy controls. Biomarkers in Medicine. 2019;13:1447-1457.
- Carro E, Bartolomé F, Bermejo-Pareja F, Villarejo-Galende A, Molina JA, Ortiz P, Calero M, Rabano A, Cantero JL, Orive G. Early diagnosis of mild cognitive impairment and Alzheimer's disease based on salivary lactoferrin. Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring. 2017;8:131-138.
- Bougea A, Koros C, Stefanis L. Salivary alpha-synuclein as a biomarker for Parkinson's disease: A systematic review. Journal of Neural Transmission. 2019;126: 1373-1382.
- Håversen L, Ohlsson BG, Hahn-Zoric M, Hanson LÅ, Mattsby-Baltzer I. Lactoferrin down-regulates the LPS-induced cytokine production in monocytic cells via NF-κB. Cellular Immunology. 2002;220:83-95.
- 84. Moreno-Navarrete JM, Ortega FJ, Bassols J, Castro A, Ricart W, Fernández-Real JM. Association of circulating lactoferrin concentration and 2 nonsynonymous LTF gene polymorphisms with dyslipidemia in men depends on glucose-tolerance status. Clinical Chemistry. 2008;54:301-309.
- 85. Moreno-Navarrete JM, Ortega FJ, Bassols J, Ricart W, Fernández-Real JM. Decreased circulating lactoferrin in insulin resistance and altered glucose tolerance as a possible marker of neutrophil dysfunction in type 2 diabetes. The Journal of Clinical Endocrinology & Metabolism. 2009;94:4036-4044.
- 86. Takeuchi T, Shimizu H, Ando K, Harada E. Bovine lactoferrin reduces plasma triacylglycerol and NEFA accompanied by decreased hepatic cholesterol and triacylglycerol contents in rodents. British Journal of Nutrition. 2004;91:533-538.
- 87. Mohamed WA, Schaalan MF. Antidiabetic efficacy of lactoferrin in type 2 diabetic

pediatrics; controlling impact on PPAR- $\gamma$ , SIRT-1, and TLR4 downstream signaling pathway. Diabetology & Metabolic Syndrome. 2018;10:1-12.

- Yang D, de la Rosa G, Tewary P, Oppenheim JJ. Alarmins link neutrophils and dendritic cells. Trends in Immunology. 2009;30:531-537.
- 89. De la Rosa G, Yang D, Tewary P, Varadhachary A, Oppenheim JJ. Lactoferrin acts as an alarmin to promote the recruitment and activation of APCs and antigen-specific immune responses. The Journal of Immunology. 2008;180:6868-6876.
- Fillebeen C, Ruchoux M-M, Mitchell V, 90. Vincent S, Benaïssa M, Pierce A. Lactoferrin is synthesized by activated microglia in the human substantia nigra and its synthesis by the human microglial CHME cell line is upregulated by tumor necrosis factor α or 1-methyl-4phenylpyridinium treatment. Molecular Brain Research. 2001;96:103-113.
- 91. Embleton ND, Berrington JE, McGuire W, Stewart CJ, Cummings SP. Lactoferrin: Antimicrobial activity and therapeutic potential. 2013;143-149.
- 92. Wada Y, Lönnerdal B. Bioactive peptides derived from human milk proteins mechanisms of action. The Journal of Nutritional Biochemistry. 2014;25:503-514.
- 93. Vogel P, Donoviel MS, Read R, Hansen GM, Hazlewood J, Anderson SJ, Sun W, Swaffield J, Oravecz T. Incomplete inhibition of sphingosine 1-phosphate lyase modulates immune system function yet prevents early lethality and non-lymphoid lesions. PloS One. 2009;4:e4112-e4112.
- 94. Ma H, Shieh K-J. ELISA Technique. Nature and Science. 2006;36.
- 95. Peen E, Almer S, Bodemar G, Ryden BO, Sjölin C, Tejle K, Skogh T. Anti-lactoferrin antibodies and other types of ANCA in

ulcerative colitis, primary sclerosing cholangitis, and Crohn's disease. Gut. 1993;34:56-62.

- 96. Ohashi Y, Ishida R, Kojima T, Goto E, Matsumoto Y, Watanabe K, Ishida N, Nakata K, Takeuchi T, Tsubota K. Abnormal protein profiles in tears with dry eye syndrome. American Journal of Ophthalmology. 2003;136:291-299.
- 97. Jensen OL, Gluud BS, Birgens HS. The concentration of lactoferrin in tears of normals and of diabetics. Acta Ophthalmologica. 1986;64:83-87.
- Yu L, Chen X, Qin G, Xie H, Lv P. Tear film function in type 2 diabetic patients with retinopathy. Ophthalmologica. 2008;222: 284-291.
- 99. Muratsu K, Morioka T. Levels of salivary lysozyme, lactoperoxidase, and lactoferrin in diabetic hamsters. Infection and Immunity. 1985;48:389-394.
- 100. Vengen IT, Dale AC, Wiseth R, Midthjell K, Videm V. Lactoferrin is a novel predictor of fatal ischemic heart disease in diabetes mellitus type 2: Long-term follow-up of the HUNT 1 study. Atherosclerosis. 2010;212: 614-620.
- EI-Desouky MA, Osman S, Shams Eldin NM, Emaraa I. Arginase enzyme activity and lactoferrin protein concentration in Egyptian diabetic patients. Int J Adv Res. 2017;5:1518-1523.
- 102. Chorzewski M, Orywal K, Sierpinska T, Golebiewska M. Salivary protective factors in patients suffering from decompensated type 2 diabetes. Advances in Medical Sciences. 2017;62:211-215.
- 103. Abdulkader RT, Kadhim NA, Muhsin FY. Lactoferrin A promising sign for developing peripheral neuropathy in patients with type 2 diabetes mellitus. Annals of the Romanian Society for Cell Biology. 2021; 359-369.

© 2021 Alhalwani; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/79892